

Evaluation of rapid immuno-chromatographic test to differentiate between mycobacterium tuberculosis complex and nontuberculous mycobacteria

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Abstract

Background: Strategies used for the clinical management of patients with Mycobacterium tuberculosis complex (MTBC) and Nontuberculous Mycobacteria (NTM) are different, therefore, prompt detection, isolation, and discrimination is necessary for suitable management. MPT64 TB Ag test was found to be a rapid immunochromatographic test for their differentiation. The aim of the present was to evaluate a commercial assay, SD TB Ag MPT64 Rapid for characterization of Mycobacteria isolated on Lowenstein Jensen (L-J) medium. **Material and Methods:** A total of 100 isolates recovered from both pulmonary (50 isolates) as well as extra pulmonary specimens (50 isolates) were characterized as MTBC or NTM based on standard phenotypic characteristics and biochemical tests. These results were compared with commercial SD TB Ag MPT64 Rapid test. **Results:** Among 100 mycobacterial isolates, 98 isolates were identified as MTBC and the two isolates from extra pulmonary cases were identified as NTM on the basis of biochemical tests. There was no discrepancy found in differentiation of these isolates between biochemical tests and TB Antigen MPT64 rapid test. **Conclusion:** The sensitivity and specificity of TB Antigen MPT64 rapid test was 100% as compared to biochemical methods. The MPT64 TB Ag test was found to be a simple, rapid and reliable test to differentiate MTBC from NTM.

Keywords: Tuberculosis, MPT 64, Mycobacterial characterization, sensitivity, specificity.

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Received Date: 18/08/2016 Revised Date: 24/09/2016 Accepted Date: 12/10/2016

Access this article online

Quick Response Code:



Website:

www.statperson.com

DOI: 14 October
2016

INTRODUCTION

Tuberculosis remains a major public health problem despite noteworthy socio-economic development, advances and availability of technology¹. The capacity of the laboratories to perform a rapid culture, identification

and differentiation of *M. tuberculosis* complex (MTBC) from non-tuberculous mycobacteria (NTM) is vital in the management of tuberculosis patients. NTM are inherently resistant to conventional anti-tuberculosis drugs, require modified treatment regimens and are often misdiagnosed as multidrug-resistant tuberculosis². Strategies used for the clinical management of patients with MTBC and NTM are different, therefore, prompt detection, isolation, and discrimination is necessary for suitable management. Conventional biochemical methods can identify mycobacterial species; but these are laborious, time-consuming and error-prone. Although molecular methods of identification are reliable, but require a specialized set up, sophisticated and expensive equipment, trained laboratory personnel and are expensive for resource-poor countries. The *M. tuberculosis* protein 64 (MPT64) is a *M. tuberculosis*

complex specific antigen secreted during the bacterial growth, and is an excellent antigen for the identification of MTBC³. Studies have revealed that the MPT 64 is specific for MTBC⁴. This study was carried out to evaluate clinical usefulness of immunochromatographic test (ICT) kit based on mouse monoclonal anti-MPT 64 for discrimination between MTBC and NTM in clinical isolates from patients with pulmonary and extrapulmonary tuberculosis.

MATERIAL AND METHODS

In this prospective study, a total of 100 mycobacterial isolates on Lowenstein-Jensen (L-J) medium recovered from both pulmonary (50 isolates) as well as extra pulmonary specimens (50 isolates) from adult patients were included. The isolates were confirmed by Ziehl - Neelsen (ZN) staining. The positive cultures were screened to differentiate the growth of mycobacteria into MTBC and NTM by niacin accumulation test, nitrate reduction test, heat stable catalase at 68°C/pH7 and Para-nitrobenzoic acid (PNB) susceptibility test⁵. The cultures which were not identified as MTBC and suspected to be NTM were further identified by rate of growth, pigment production, Urease test, Tween – 80 hydrolysis test, Arylsulfatase test, Mac Conkey agar test and Sodium Chloride tolerance test⁵. All the cultures were also subjected to TB Antigen MPT64 rapid test for differentiation into MTBC and NTM. Briefly, 3-4 colonies from L-J were emulsified in 200 µl of extraction buffer. 100 µl of suspended solid culture in buffer was added into the sample well. The inoculated cassettes were kept undisturbed at room temperature and were examined at the end of 15 minutes for presence of pink band in “Control” and “Test” region⁶. The appearance of control band confirmed the validity of the test. If the control band was not visible in 15 minutes, the result was considered invalid and the sample was retested. The presence of only control band in the absence of test band was considered a negative test. Presence of both control and test band indicated a positive test.

RESULTS

One hundred mycobacterial isolates on L-J media were included in the present study and demographic details of these patients were recorded. Maximum number of patients belonged to 16- 45 years (83%) of age group followed by 46-60 years (16%) of age group in both pulmonary and extrapulmonary cases. There were 53 males and 47 females with male predominance in extra pulmonary cases (M:F=1.27:1). The pus (60%) was most common specimen followed by pleural fluid (14%) and lymph node aspirate (12%) amongst extrapulmonary specimens. Pleural involvement (32%; pleural effusion

and empyema thoracis) followed by lymph node involvement (18%) was most commonly seen in extrapulmonary cases. Other sites (16%) included axillary, mastoid and submandibular abscesses. Most common symptom in pulmonary cases was cough (92%) followed by fever (74%) and weight loss (64%) while in extrapulmonary cases most common symptom was fever (44%) followed by lymphadenopathy (36%). Out of 50 culture positive pulmonary specimens, only 23 (46%) were positive on primary ZN smear. Whereas, among 50 culture positive extrapulmonary specimens, only 18 (36%) were primary ZN smear positive. All the isolates from pulmonary cases (n=50) were positive for niacin and nitrate reduction test while negative for catalase and PNB test. In extrapulmonary isolates, 48 were positive for niacin test, negative for catalase and PNB test while 49 were positive for nitrate reduction test. The two isolates from extrapulmonary cases were positive for PNB test. Therefore, all the 98 isolates were identified as MTBC and the two isolates from extrapulmonary cases identified as NTM on the basis of biochemical tests. These two NTM isolates from extrapulmonary specimens were further identified as *Mycobacterium fortuitum* and *Mycobacterium chelonae* on the basis of biochemical tests. These isolates belong to Group IV (Rapid Growers) of Runyon's Classification. When subjected to TB Ag MPT64 Rapid test, the control band was seen in all the tested cultures (n=100), validating the test. All the 50 isolates from pulmonary cases and 48 isolates from extrapulmonary cases showed positive results (visible band for MPT64 antigen) and the two isolates from extrapulmonary cases gave negative results. There was no discrepancy found in differentiation of these isolates between biochemical tests and TB Antigen MPT64 rapid test. The sensitivity and specificity of TB Antigen MPT64 rapid test was found to be 100% as compared to biochemical methods.

DISCUSSION

In the present study, majority of patients (83%) belonged to 16 – 45 years of age group in both pulmonary and extrapulmonary cases with slight male predominance (M:F = 1.27:1) seen in extrapulmonary cases. Similar demographic profile of patients was found in a study by Sivasankari et al⁷. Pus (60%) was most common specimen and lymph node aspirate (12%) amongst extrapulmonary specimens in the present study. Previous studies also found pus as the most common specimen followed by lymph node⁸⁻¹². In the present study, most common symptom in pulmonary cases was cough (92%) followed by fever (74%) and weight loss (64%) while extrapulmonary cases showed fever (44%) as most common symptom followed by lymphadenopathy

(36%). Sharma *et al*¹³ also reported fever (98%) and cough (95%) as most common symptom in pulmonary tuberculosis. Whereas Koshti *et al*¹⁴ documented cough (66.6%) as most common symptom in pulmonary cases while fever (86.8%) followed by weight loss (58.1%) in extra-pulmonary cases. In the present study, 46% and 36% of Z-N smear positivity was seen in pulmonary and extrapulmonary cases respectively. A study published by Selvakumar *et al*, reported ZN smear sensitivity of 47% from liquefied sputum samples¹⁵. It was found that the sensitivity of extra-pulmonary smears was low as compared to the smears from pulmonary specimens. Paucibacillary nature of disease, inadequate sample or too long storage of sample may result in low smear positivity in extra pulmonary specimens. In the present study, 98% isolates were found *M.tuberculosis* complex while only 2% were NTM. Both the NTM were isolated from extrapulmonary tuberculosis cases. There was no NTM isolated from pulmonary tuberculosis cases. MTBC was found to be the most common isolate than NTM in previous studies. In developing countries like India, more than 90% of tuberculosis infections are still caused by *M. tuberculosis*. In a scenario of NTM disease prevalence and increasing drug resistance in MTB, speciation and drug susceptibility testing have become a necessity today for appropriate patient management. There is a need for a rapid, simple yet accurate test for differentiation of mycobacteria. The major attraction for using MPT64 TB Ag test was its claim to characterize mycobacterial isolates accurately in 15 minutes with a low cost (Rs. 125/- per test). This assay was evaluated for rapid characterization of 100 culture isolates and whether it could replace the tedious conventional phenotypic methods. An important observation of this study was 100% specificity and sensitivity for the assay. Other studies have also demonstrated specificity of 100% and sensitivity ranging from 96.5% to 100%^{4,16,17}. To conclude, TB Antigen MPT64 test was found to be rapid, easy to perform and cost effective with high sensitivity and specificity to identify MTBC from culture isolates. It is It can also differentiate MTBC from NTM.

REFERENCES

1. TB India 2016 RNTCP. Annual Status Report. New Delhi: Central TB Division. Directorate General of Health Services, Ministry of Health and Family Welfare. Available from: www.tbcindia.org.
2. Wagner D, Young L. Nontuberculous mycobacterial infections: A clinical review. *Infection* 2004; 32; 257-70.
3. Andersen P, Askgaard D, Ljingqvist L, et al. Proteins released from *Mycobacterium tuberculosis* during growth. *Infect Immun* 1991; 59: 1905-10.
4. A Chiyoji, Hirano K, Tomiyama T. Simple and Rapid identification of the *Mycobacterium tuberculosis* Complex by Immunochromatographic Assay Using Anti-MPT64 Monoclonal Antibodies. *J Clin Microbiol* 1999; 3693-7.
5. Global Tuberculosis Programme, World Health Organization Laboratory services in tuberculosis control Part III Culture; WHO/TB/98.258 1998.
6. SD TB Antigen MPT64 Rapid test; Standard Diagnostics, Inc. Kyonggi-do, Korea. <http://www.standardia.com>
7. Sivasankari P, Khyriem AB, Venkatesh K, Parija SC. Atypical Mycobacterial Infection Among HIV Seronegative Patients in Pondicherry. *Ind J Chest Dis Allied Sci* 2006; 48:107-9.
8. Dharmshale S, Bhardwaj R, Gohil A, Choudhary A. Extra-pulmonary tuberculosis in HIV patients in a tertiary care hospital, Mumbai. *Ind J Basic Applied Med Res* 2012; 1(3):205-8.
9. Gopal R, Padmavathy BK, Vasanthi S, Jayashree K. Extra-pulmonary tuberculosis- A retrospective study. *Ind J Tuberc* 2001; 48:225.
10. Paramasivan CN, Kumar V, Alexander C, Venkatesan P. Use of multiple media for the cultivation of mycobacteria from specimens other than sputum. *Ind J Med Res* 1987; 290-4.
11. Arora VK, Gupta R. Trends of extrapulmonary tuberculosis under Revised National Tuberculosis Control Programme: A study from South Delhi. *Ind J Tuberc* 2005.
12. Sharma SK, Mohan A. Extra pulmonary Tuberculosis. *Ind J Med Res* 2004; 120:316-53.
13. Sharma V, Sharma Y, Joshi D, Shah A. Prevalence of HIV positivity in extrapulmonary tuberculosis patients at a tertiary care hospital in central Gujarat : A Retrospective Study. *Ind J Resp Med* 2013; 2(2); 57-62.
14. Koshti A, Shrivastava A, Kapoor N. Pattern of clinical manifestation of HIV in patients of tuberculosis diagnosed by different diagnostic modalities; *Int J Pharma and Bio Sci* 2013; 4(2): (B)472 -7.
15. Selvakumar N, Sekar MG, Kumar V, Rao DVB, Rahman F, Narayanan PR. Sensitivity of Ziehl- Neelsen method for centrifuged deposit samples transported in cetylpyridinium chloride. *Ind J Med Res* 2006; 124:439-42.
16. Kumar VG, Urs TA, Ranganath RR. MPT 64 antigen detection for rapid confirmation of *M. tuberculosis* isolates. *BMC Res Notes*. 2011; 4:79.
17. Tohir AOS, Rasolofo V, Andrianarisoa SH, Ranjalaly GM, Ramarokoto H. Validation of an immunochromatographic assay kit for the identification of the *Mycobacterium tuberculosis* complex; *Mem Inst Oswaldo Cruz, Rio de Janeiro* 2011; 106(6): 777-80.

Source of Support: None Declared
Conflict of Interest: None Declared