

Characterization of rho-associated protein kinase Rock1 by using online bioinformatics tools

Arun S Kharat¹, Ashish B Gulwe^{2*}

¹Director, UGC, Academic Staff College, Dr. Babasaheb Ambedkar Marathwada University, Aurangabad, Maharashtra, INDIA.

²Assistant Professor, Swami Ramanand Teerth Marathwada University Nanded, Sub Centre, School of Technology, Latur-413512, Maharashtra, INDIA.

Email: ashishgulwe@gmail.com

Abstract

Abstract: ROCK-signalling system is involved in regulation of cellular growth, and life cycle through control of muscle cell contractility, change in ROCK pathway and resultant impaired cell contractility cause to disease in different organs, including cardiovascular, respiratory and renal systems. Hence, ROCK inhibitors are potential therapeutic agents for hypertension, heart disease, pulmonary disease, asthma, erectile dysfunction, diabetic renal failure, chronic nephritis and glaucoma. ROCK inhibitors, similar to some other cytoskeletal drugs, could increase matrix metalloproteinase expression in cells and may induce extracellular matrix reorganization, ROCK inhibitors could weaken cell attachment to its extracellular matrix, which results in relaxation of the whole of tissue and hence, it is also probable that ROCK inhibitors enhance outflow through unknown mechanisms. Rho-associated kinase inhibitors relax smooth muscle tone in brain vasculature and could potentially increase optic nerve head perfusion. ROCK inhibitors could have neuroprotective effects on ganglion cells.

Keywords: Molecular function Kinase, Serine/threonine-protein kinase, Transferase matrix assembly process, essential for osteoblast mineralization.

*Address for Correspondence:

Mr. Ashish B Gulwe, Assistant Professor, Swami Ramanand Teerth Marathwada University Nanded, Sub Centre, School of Technology, Latur-413512, Maharashtra, INDIA.

Email: ashishgulwe@gmail.com

Received Date: 09/02/2015 Revised Date: 18/02/2015 Accepted Date: 22/02/2015

Access this article online	
Quick Response Code:	Website: www.statperson.com
	DOI: 23 February 2015

INTRODUCTION

Protein kinase which is a key regulator of actin cytoskeleton and cell polarity. Involved in regulation of

smooth muscle contraction, actin cytoskeleton organization, stress fiber and focal adhesion formation, neurite retraction, cell adhesion and motility. Acts as a suppressor of inflammatory cell migration by regulating. Required for centrosome positioning and centrosome-dependent exit from mitosis. Plays a role in terminal erythroid differentiation. May regulate closure of the eyelids and ventral body wall by inducing the assembly of actomyosin bundles. Promotes keratinocyte terminal differentiation. Involved in osteoblast compaction.

Annotation

ROCK1 Gene Structure

Chromosome: chr18

Orientation: -

Length coding sequence: 4062 nucleotides.

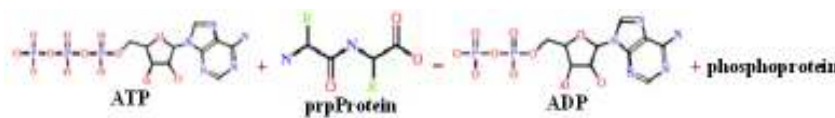
Table 1

Region	Start	End	Region Length	Phase at end
UTR	18,690,872	18,691,812		
Exon	18,690,779	18,690,871	93	0
Exon	18,650,493	18,650,574	82	1
Exon	18,629,741	18,629,841	101	0
Exon	18,629,053	18,629,190	138	0
Exon	18,625,253	18,625,428	176	2
Exon	18,624,063	18,624,147	85	0
Exon	18,622,526	18,622,670	145	1
Exon	18,622,058	18,622,196	139	2
Exon	18,619,433	18,619,524	92	1
Exon	18,608,737	18,608,896	160	2
Exon	18,603,581	18,603,641	61	0
Exon	18,600,112	18,600,200	89	2
Exon	18,595,392	18,595,440	49	0
Exon	18,588,020	18,588,155	136	1
Exon	18,586,660	18,586,751	92	0
Exon	18,586,312	18,586,558	247	1
Exon	18,572,792	18,572,898	107	0
Exon	18,571,137	18,571,287	151	1
Exon	18,566,911	18,567,071	161	0
Exon	18,564,312	18,564,496	185	2
Exon	18,562,724	18,562,793	70	0
Exon	18,559,871	18,559,965	95	2
Exon	18,550,309	18,550,474	166	0
Exon	18,549,076	18,549,169	94	1
Exon	18,548,733	18,548,821	89	0
Exon	18,547,713	18,547,901	189	0
Exon	18,546,878	18,547,037	160	1
Exon	18,540,097	18,540,167	71	0
Exon	18,539,801	18,539,889	89	2
Exon	18,535,128	18,535,206	79	0
Exon	18,534,744	18,535,005	262	1
Exon	18,533,539	18,533,746	208	2
Exon	18,531,345	18,531,348	4	0
UTR	18,529,703	18,531,344		

Catalytic activity: ATP + a protein = ADP + a phosphoprotein.

Enzyme reactions

Reaction: ATP + a protein = ADP + a phosphoprotein



Enzyme regulation: Activated by RHOA binding.
 Inhibited by Y-27632. Binding site 105 Active site 198
 Nucleotide binding- 82 90 Zink finger -1228 1281.
 Taxonomic lineage of the protein is as follows
 Eukaryota › Metazoa › Chordata › Craniata › Vertebrata ›
 Euteleostomi › Mammalia › Eutheria › Euarchontoglires ›
 Primates › Haplorrhini › Catarrhini › Hominidae › Homo

Structural alignment of the protein

Protein namesi Recommended name:
 Rho-associated protein kinase 1 (EC: 2.7.11.1)

Alternative name(s):
 Renal carcinoma antigen NY-REN-35
 Rho-associated, coiled-coil-containing protein kinase 1
 Rho-associated, coiled-coil-containing protein kinase I
 Short name: ROCK-I p160 ROCK-1
 Short name: p160ROCK
 Gene namesi Name: ROCK1
 Organismi Homo sapiens (Human)
 Taxonomic identifieri 9606 [NCBI]

Structural comparison of the protein

Chromosome 18PDB ID	Chain ID	Protein Name	Pfam ID	SCOP ID	UniProt ID	Z-Score
3o0z	C	Rho-associated protein kinase 1	PF02185		Q13464	4.6
2c08	A	Sh3-containing grb2-like protein 2	PF03114	a.238.1.1	O35179	2.84
2fxm	A	Myosin heavy chain, cardiac muscle beta isoform		h.1.26.1		2.73
2oqq	B	Transcription factor hy5	PF00170		O24646	2.65
2v66	B	Nuclear distribution protein nude-like 1	PF04880		Q9GZM8	2.6
2pih	A	Protein ymca	PF06133	a.281.1.1	O31779	2.58
3ssu	A	Vimentin	PF00038 PF04732		P08670	2.55
1s2x	A	Cag-z	PF09053	a.47.3.1	Q9JMX9	2.54
2v71	A	Nuclear distribution protein nude-like 1	PF04880		Q78PB6	2.53
1joc	A	Early endosomal autoantigen 1	PF01363	g.50.1.1 h.1.21.1	Q15075	2.53
3swk	A	Vimentin	PF00038		P08670	2.51
1m1j	A	Fibrinogen alpha subunit	PF00147 PF08702 PF12160	h.1.8.1	P14448	2.5
3ni0	A	Bone marrow stromal antigen 2				2.5
2f9d	P	Splicing factor 3b subunit 1	PF08920		O75533	2.47
2ykt	A	Brain-specific angiogenesis inhibitor 1-associated protein 2	PF08397		Q9UQB8	2.45
1r8i	A	Trac	PF07996	a.8.7.1	Q9L6G5	2.45

Taxonomic lineage

Eukaryota › Metazoa › Chordata › Craniata › Vertebrata › Euteleostomi › Mammalia › Eutheria › Euarchontoglires › Primates › Haplorrhini › Catarrhini › Hominidae › Homo

Secondary Structure prediction by DSSP

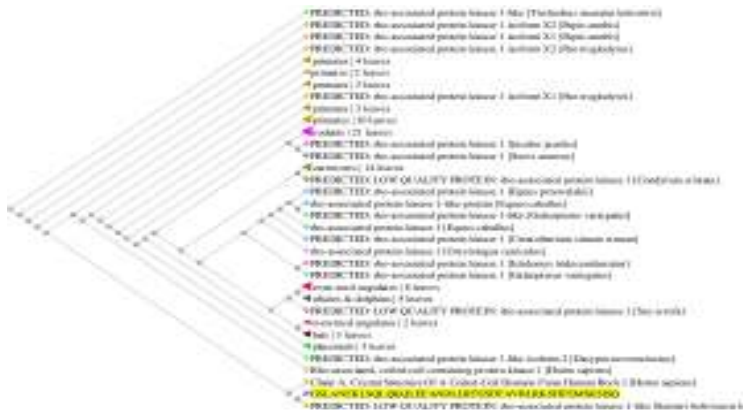
89% helical (2 helices; 150 residues)

Fasta format of the amino acid sequence

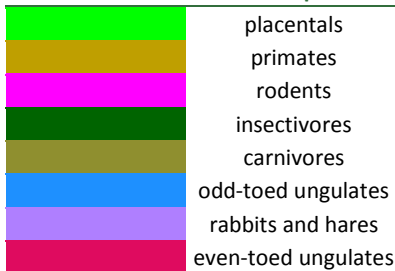
>3O0Z:D|PDBID|CHAIN|SEQUENCE

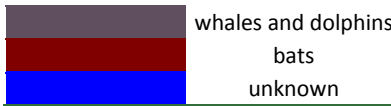
```
GSLANEKLSQLQKQLEEANDLLRTESTDAVRLRKS
HTEMKSKISQLESLNRELQERNRILENSKSTDKDY
YQLQAILEAERRDRGHDSEMIGDLQARITSLQEEV
KHLKHNLEKVEGERKEAQDMLNHSEKEKNNLEID
LNYKLKSLQQRLEQEVNEHKVTKARLTD
```

Phylogenetic Tree of the protein



Blast names color map





Tree Method: Fast Minimum Evolution

Max Seq Difference: 0.85

Distance: Grishin

Tree is based on COBALT multiple alignment. This gene encodes a protein serine/threonine kinase that is activated when bound to the GTP-bound form of Rho. The small GTPase Rho regulates formation of focal adhesions and stress fibers of fibroblasts, as well as adhesion and aggregation of platelets and lymphocytes by shuttling between the inactive GDP-bound form and the active GTP-bound form. Rho is also essential in cytokinesis and plays a role in transcriptional activation by serum response factor. This protein, a downstream effector of Rho, phosphorylates and activates LIM kinase, which in turn, phosphorylates cofilin, inhibiting its actin-depolymerizing activity. [provided by RefSeq, Jul 2008]

Function: EC: non-specific serine/threonine protein kinase; Transferases; Transferring phosphorous-containing groups; Protein-serine/threonine kinases (2.7.11.1)

Ramachandran Plot: Ramachandran Plot Analysis

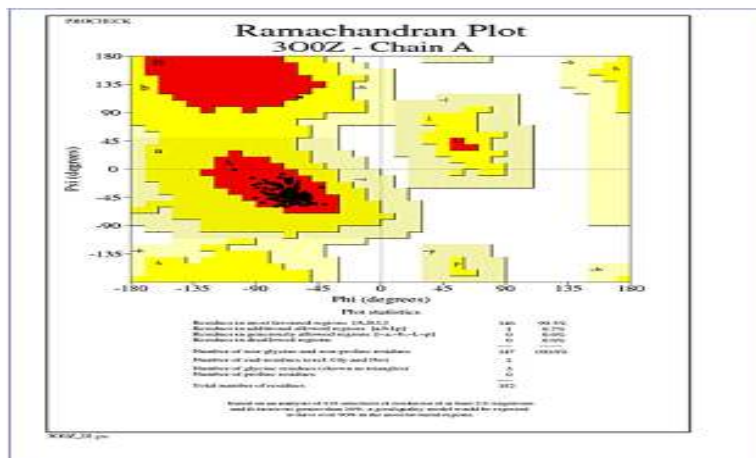
The number of residues in favoured region are 98.9%

Evaluation of residues

- Residue [A 617 :ARG] (-42.79, -28.84) in Allowed region
- Residue [B 580 :SER] (-57.27, -69.54) in Allowed region
- Residue [B 688 :VAL] (-44.48, -29.66) in Allowed region
- Residue [C 537 :ASN] (-42.95, -62.42) in Allowed region
- Residue [C 581 :LEU] (-41.31, -35.70) in Allowed region
- Residue [C 595 :SER] (-35.69, -55.80) in Allowed region
- Residue [D 586 :GLN] (-55.62, -13.14) in Allowed region
- Number of residues in favoured region (~98.0% expected): 608 (98.9%)

Number of residues in allowed region (~2.0% expected): 7 (1.1%)

Number of residues in outlier region: 0 (0.0%)



REFERENCES

1. Quigley HA, Broman AT. The number of people with glaucoma worldwide in 2010 and 2020. *Br J Ophthalmol* 2006;90:262-267.
2. Marquis RE, Whitson JT. Management of glaucoma: Focus on pharmacological therapy. *Drugs Aging* 2005; 22:1-21.
3. Gordon MO, Beiser JA, Brandt JD, Heuer DK, Higginbotham EJ, Johnson CA, *et al.* The ocular hypertension treatment study: Baseline factors that predict the onset of primary open-angle glaucoma. *Arch Ophthalmol* 2002; 120:714-720.
4. Kass MA, Heuer DK, Higginbotham EJ, Johnson CA, Keltner JL, Miller JP, *et al.* The Ocular Hypertension Treatment Study: A randomized trial determines that topical ocular hypotensive medication delays or prevents the onset of primary open-angle glaucoma. *Arch Ophthalmol* 2002; 120:701-713.
5. Heijl A, Leske MC, Bengtsson B, Hyman L, Bengtsson B, Hussein M, *et al.* Reduction of intraocular pressure and glaucoma progression: Results from the Early Manifest Glaucoma Trial. *Arch Ophthalmol* 2002; 120:1268-1279.
6. Leske MC, Heijl A, Hyman L, Bengtsson B, Dong L, Yang Z, *et al.* Predictors of long-term progression in the

- early manifest glaucoma trial. *Ophthalmology* 2007; 114:1965-1972.
7. Francis BA, Alvarado J. The cellular basis of aqueous outflow regulation. *Curr Opin Ophthalmol* 1997; 8:19-27.
 8. Rao VP, Epstein DL. Rho GTPase/Rho kinase inhibition as a novel target for the treatment of glaucoma. *BioDrugs* 2007; 21:167-177.
 9. Gabelt BT, Kaufman PL. Changes in aqueous humor dynamics with age and glaucoma. *Prog Retin Eye Res* 2005; 24:612-637.
 10. Lütjen-Drecoll E. Functional morphology of the trabecular meshwork in primate eyes. *Prog Retin Eye Res* 1999; 18:91-119.
 11. Tripathi RC, Li J, Chan WF, Tripathi BJ. Aqueous humor in glaucomatous eyes contains an increased level of TGF-beta 2. *Exp Eye Res* 1994; 59:723-727.
 12. Wiederholt M, Thieme H, Stumpff F. The regulation of trabecular meshwork and ciliary muscle contractility. *Prog Retin Eye Res* 2000; 19:271-295.
 13. Yorio T, Krishnamoorthy R, Prasanna G. Endothelin: Is it a contributor to glaucoma pathophysiology? *J Glaucoma* 2002; 11:259-270.
 14. Tezel G, Kass MA, Kolker AE, Becker B, Wax MB. Plasma and aqueous humor endothelin levels in primary open-angle glaucoma. *J Glaucoma* 1997; 6:83-89.
 15. Lütjen-Drecoll E. Morphological changes in glaucomatous eyes and the role of TGFbeta2 for the pathogenesis of the disease. *Exp Eye Res* 2005; 81:1-4.
 16. Challa P, Arnold JJ. Rho-kinase inhibitors offer a new approach in the treatment of glaucoma. *Expert Opin Investig Drugs* 2014; 23:81-95.
 17. Wang H, Cheng JW, Wei RL, Cai JP, Li Y, Ma XY. Meta-analysis of selective laser trabeculoplasty with argon laser trabeculoplasty in the treatment of open-angle glaucoma. *Can J Ophthalmol* 2013; 48:186-192.
 18. Brusini P. Canaloplasty in open-angle glaucoma surgery: A four-year follow-up. *ScientificWorldJournal* 2014; 2014:469609.
 19. Brandão LM, Grieshaber MC. Update on minimally invasive glaucoma surgery (MIGS) and new implants. *J Ophthalmol* 2013; 2013:705915.
 20. Voskanyan L, Garcia-Feijó J, Belda JI, Fea A, Jünemann A, Baudouin C, *et al.* Prospective, unmasked evaluation of the iStent® inject system for open-angle glaucoma: Synergy trial. *Adv Ther* 2014; 31:189-201.
 21. Hays CL, Gulati V, Fan S, Samuelson TW, Ahmed II, Toris CB. Improvement in outflow facility by two novel microinvasive glaucoma surgery implants. *Invest Ophthalmol Vis Sci* 2014; 55:1893-1900.
 22. Jordan JF, Wecker T, van Oterendorp C, Anton A, Reinhard T, Boehringer D, *et al.* Trabectome surgery for primary and secondary open angle glaucomas. *Graefes Arch Clin Exp Ophthalmol* 2013; 251:2753-2760.
 23. Wilmsmeyer S, Philippin H, Funk J. Excimer laser trabeculotomy: A new, minimally invasive procedure for patients with glaucoma. *Graefes Arch Clin Exp Ophthalmol* 2006; 244:670-676.
 24. Inoue T, Tanihara H. Rho-associated kinase inhibitors: A novel glaucoma therapy. *Prog Retin Eye Res* 2013; 37:1-12.
 25. Somlyo AP, Somlyo AV. Ca²⁺ sensitivity of smooth muscle and nonmuscle myosin II: Modulated by G proteins, kinases, and myosin phosphatase. *Physiol Rev* 2003; 83:1325-1358.
 26. Fukata Y, Amano M, Kaibuchi K. Rho-Rho-kinase pathway in smooth muscle contraction and cytoskeletal reorganization of non-muscle cells. *Trends Pharmacol Sci* 2001; 22:32-39.
 27. WettSchureck N, Offermanns S. Rho/Rho-kinase mediated signaling in physiology and pathophysiology. *J Mol Med (Berl)* 2002; 80:629-638.
 28. Amano M, Chihara K, Kimura K, Fukata Y, Nakamura N, Matsuura Y, *et al.* Formation of actin stress fibers and focal adhesions enhanced by Rho-kinase. *Science* 1997; 275:1308-1311.
 29. Uehata M, Ishizaki T, Satoh H, Ono T, Kawahara T, Morishita T, *et al.* Calcium sensitization of smooth muscle mediated by a Rho-associated protein kinase in hypertension. *Nature* 1997; 389:990-994.
 30. Rao MY, Soliman H, Bankar G, Lin G, MacLeod KM. Contribution of Rho kinase to blood pressure elevation and vasoconstrictor responsiveness in type 2 diabetic Goto-Kakizaki rats. *J Hypertens* 2013; 31:1160-1169.
 31. Li Y, Zhu W, Tao J, Xin P, Liu M, Li J, *et al.* Fasudil protects the heart against ischemia-reperfusion injury by attenuating endoplasmic reticulum stress and modulating SERCA activity: The differential role for PI3K/Akt and JAK2/STAT3 signaling pathways. *PLoS One* 2012; 7:e48115.
 32. Li Q, Xu Y, Li X, Guo Y, Liu G. Inhibition of Rho-kinase ameliorates myocardial remodeling and fibrosis in pressure overload and myocardial infarction: Role of TGF-β1-TAK1. *Toxicol Lett* 2012; 211:91-97.
 33. S.C. Lovell, I.W. Davis, W.B. Arendall III, P.I.W. de Bakker, J.M. Word, M.G. Prisant, J.S. Richardson and D.C. Richardson (2002) Structure validation by Calpha geometry: phi,psi and Cbeta deviation. *Proteins: Structure, Function and Genetics.* 50: 437-450.
 34. Depth: a web server to compute depth, cavity sizes, detect potential small-molecule ligand-binding cavities and predict the pKa of ionizable residues in proteins Kuan Pern Tan, Thanh Binh Nguyen, Siddharth Patel, Raghavan Varadarajan and M. S. Madhusudhan *Nucl. Acids Res.* (1 July 2013) 41 (W1): W314-W321. doi: 10.1093/nar/gkt503

Source of Support: None Declared
Conflict of Interest: None Declared